## Gateway LR recombination

Janes Lab Protocols

- 1. In a microcentrifuge tube, add 150 ng entry clone + 150 ng destination vector in 8  $\mu$ l TE buffer.
- 2. Remove the LR Clonase<sup>™</sup> II Enzyme Mix (Invitrogen #11791-020) from –20° C and thaw on ice (~2 minutes).
- 3. Vortex the LR Clonase<sup>™</sup> II Enzyme Mix briefly twice (2 seconds each time).
- 4. Add 2 μl of LR Clonase™ II Enzyme Mix to each sample. Mix well by vortexing briefly twice (2 seconds each time).
  - Return LR Clonase™ II Enzyme Mix to –20°C immediately after use
- 5. Incubate the reactions at room temperature. Take 5 μl of the sample after 1 hr and proceed to Step 6. Leave the remaining 5 μl of each sample at room temperature for up to 18 hr.
- 6. Add 0.5 µl of the Proteinase K solution to the 5 µl reaction and incubate for 10 minutes at 37°C.
  Proteinase K cleaves and inactivates the LR Clonase™ II Enzyme Mix
- 7. Ethanol precipitate the reaction and resuspend in 5 µl water.
- 8. Transform 1 µl of the resuspended reaction into electrocompetent bacteria (see Janes PCRcloning.pdf).
- 9. If the transformation is unsuccessful, repeat Steps 6–8 with the 18-hr recombination fraction.