# Sample preparation

## Learning objectives

 Understand the differences in sample preparation for different sample formats

 Appreciate the critical steps for embedding and cryosectioning of tissue samples for LCM

 Embed, section, and stain representative tissues and adherent cells

## Three main sample preps

Suspension cells: isolate and lyse

Cultured adherent cells: fix, stain, and microdissect

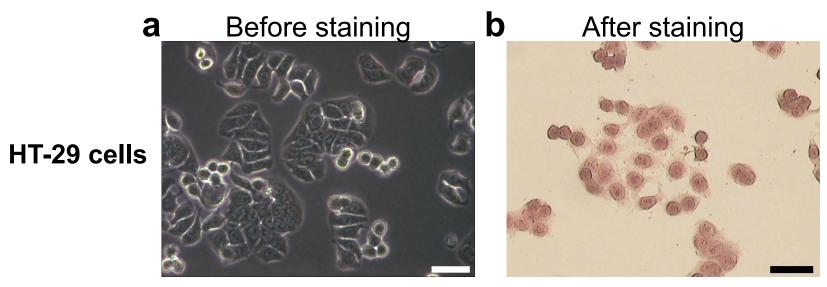
 Tissues: embed, cryosection, fix, stain, and microdissect

## Suspension preparations

- Add 0.1% (w/v) saponin to the first step of poly(A) amplification
- Permeabilize cells and release mRNAs
- Does not interfere with downstream enzymatic steps

#### Cultured adherent cells

- EtOH fixation on glass
- Histologic staining (nuclear fast red, hematoxylin, toluidine blue) < 2 min aqueous!</li>
- EtOH dehydration and xylene clearing

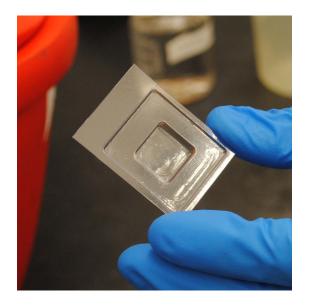


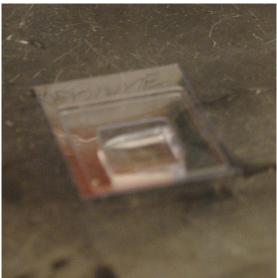
NFR = brownish red

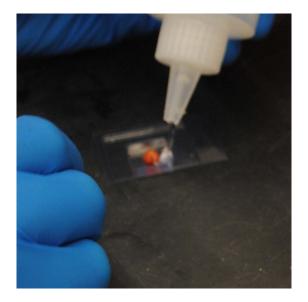
### Tissue embedding

- Snap freeze tissue pieces <u>immediately</u> in liquid nitrogen or dry ice-isopentane (store at −80°C)
- Embed in a small cryomold with NEG50 (like OCT but less flaky at low temps)—make sure that the tissue piece is completely covered
- Work quickly and accept some thawing on the exterior of the tissue piece (do not sample on the edge)
- Blocks can be stored long term, but sectioned ONCE

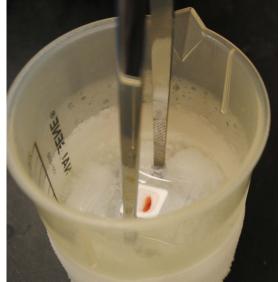
# Embedding workflow

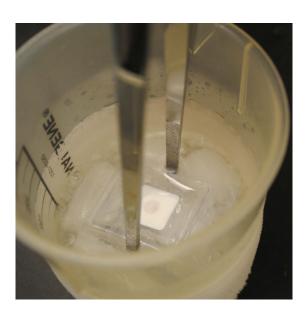




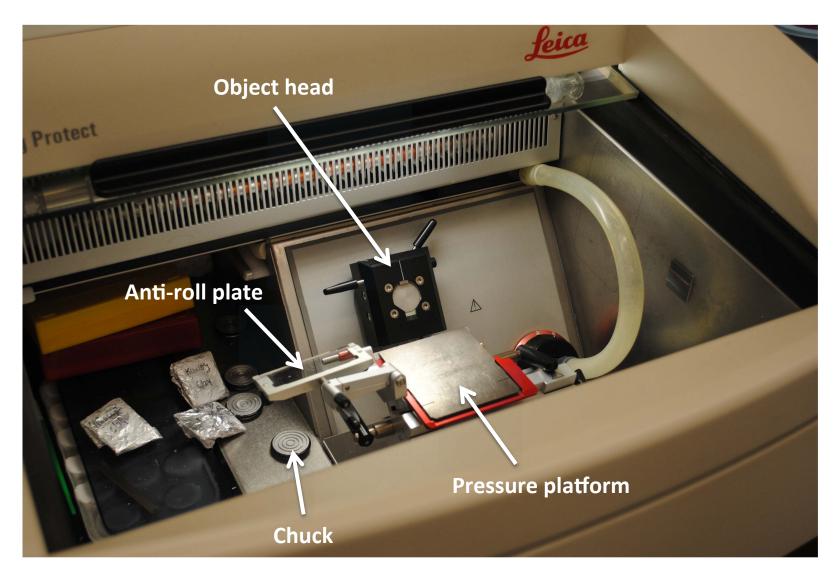






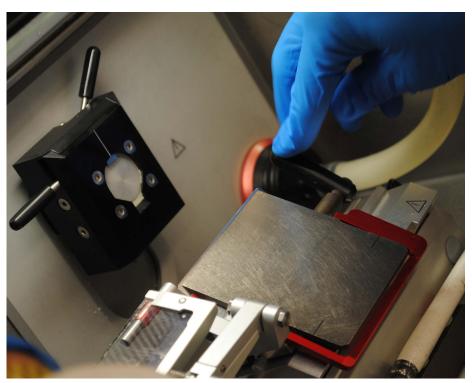


## Cryostat nomenclature



### Before starting

• Equilibrate at a cold temperature (< −20°C)

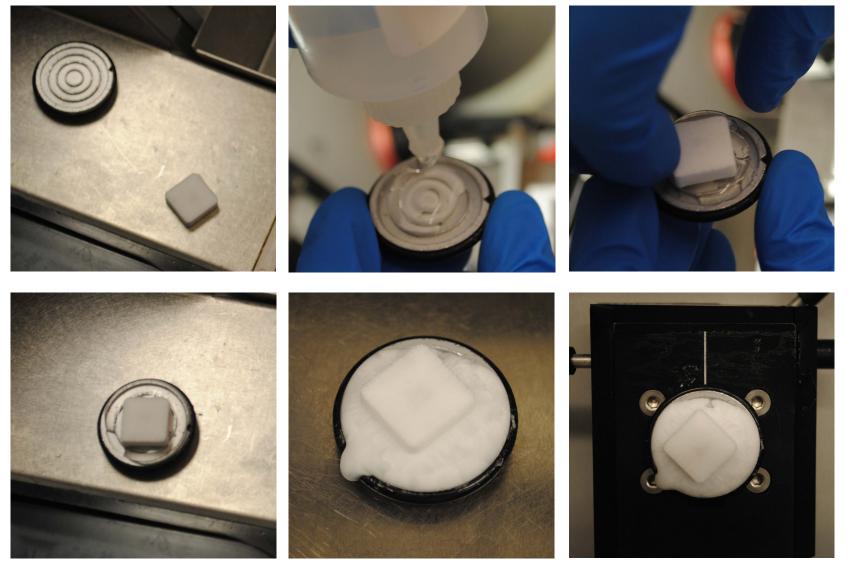


Fresh blade



Moderate platform angle and move platform to object head

## Setting up the cryoblock



Diagonal (to reduce friction)

#### Preserving genetically encoded fluorescence

- Proteins diffuse much more rapidly than RNAs upon sectioning
- Soak snap-frozen tissue in 100% EtOH at −80°C for 24 hrs
- Transfer to a fresh conical tube at –80°C and vent daily for ~1 week
- Embed and section as for any other tissue

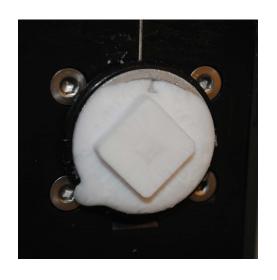
Before LCM After LCM LCM cap

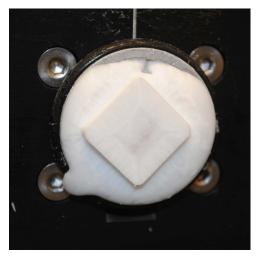
TdT-labeled OPCs

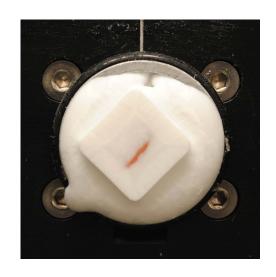
After LCM

LCM cap

#### Trim, trim, trim... section









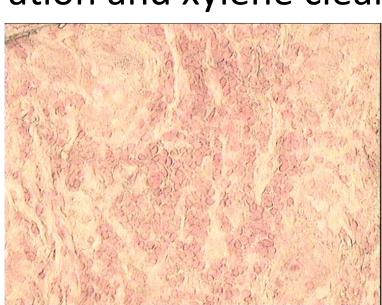
Refreeze sections quickly inside the cryostat box! (Precool slides if needed.)

Try untreated slides and "Plus" slides depending on the application

## Cryosection storage and staining

- Transport slides to –80°C on dry ice
- EtOH fix slides at the −80°C (no thawing)
- Histologic staining (nuclear fast red, hematoxylin, toluidine blue) < 2 min aqueous!</li>
- EtOH dehydration and xylene clearing

Human melanoma



## Questions?